

Human High sensitivity IFN γ ELISA KIT

INTENDED USE

The Human high sensitivity IFN γ ELISA is to be used for the in-vitro quantitative determination interferon gamma (IFN γ) in human sera, plasmas, buffered solutions or cell culture media. The assay will recognize both natural and recombinant human IFN γ . **This kit is intended for research use only and is not to be used in diagnostic procedures.**

PRINCIPLE OF THE METHOD

The IFN γ Kit is a solid phase sandwich Enzyme Linked-Immuno-Sorbent Assay (ELISA). A monoclonal antibody specific for IFN γ has been coated onto the wells of the microtiter strips provided. Samples, including standards of known IFN γ concentrations and unknowns are pipetted into these wells.

During the first incubation, the IFN γ antigen and a biotinylated monoclonal antibody specific for IFN γ are successively incubated. After washing, the enzyme (streptavidin-peroxydase) is added. All the unbound enzyme is removed by washing and the first amplification step is performed by adding the Biotine-Tyramine reagent. Under the action of HRP, a biotine polymerisation reaction occurs in the region of the HRP linked to the detection antibody. After washing the second amplification step is performed and the polymerised biotine is revealed by a new streptavidin-HRP step. Finally after washing, the substrate is added. The intensity of this coloured product is directly proportional to the concentration of IFN γ present in the samples.

REAGENTS PROVIDED AND RECONSTITUTION

REAGENTS (Store at 2-8°C)	COLOR CODE	1x96 wells Cat # 850.900.096	2x96 wells Cat # 850.900.192	RECONSTITUTION
96-wells microtiter plates	-	1	2	Ready-to-use.
Plastic cover	-	2	4	-
Standard : 25 pg/ml	Yellow	2 vials	4 vials	Reconstitute with the volume of standard diluent indicated on the vial. (See Reagents Preparation).
Standard Diluent Buffer	Black	1 vial	1 vial	(25 ml) 10X concentrate. Dilute in distilled Water.
Biotinylated anti-IFN γ	Red	1 vial	2 vials	(0.4 ml) Dilute in biotinylated antibody diluent.
Biotinylated Antibody Diluent	Red	1 vial (7.5 ml)	1 vial (13 ml)	Ready-to-use.
Streptavidin-HRP		2 vials	4 vials	(5 μ l) – Add 0.5ml of HRP-Diluent before further dilutions
Amplification Diluent	Brown & blue spot	1 vial	1 vial	(25 ml) Ready to use.
Amplifier*	Yellow	1 vial	2 vials	(200 μ l) Dilute in Amplification buffer.
HRP Diluent	Red	1 vial	2 vials	(25 ml) Ready-to-use
Washing Buffer	White	1 vial	2 vials	(10 ml) 200X concentrate. Dilute in distilled Water.
Chromogen TMB :	Brown	1 vial (11 ml)	1 vial (24 ml)	Ready-to-use.
H ₂ SO ₄ : Stop Reagent	Black	1 vial	2 vials	(11 ml) Ready-to-use.

*Reagent contains ethyl alcohol.

MATERIAL REQUIRED BUT NOT PROVIDED

- Distilled water.
- Pipettes : 10 μ l, 50 μ l, 100 μ l, 200 μ l and 1000 μ l.
- Vortex mixer and magnetic stirrer.
- Rotator set

SAFETY

- For research use only.
- No human blood components included in this kit. Nevertheless, handling of human blood derivative samples should be in accordance with local safety procedures, e.g. CDC/NIH Health manual : "Biosafety in Microbiological and Biomedical Laboratories 4th Edition" 1999"
- Avoid any skin contact with H₂SO₄ and TMB. In case of contact, wash thoroughly.
- Do not eat, drink, smoke or apply cosmetics where kit reagents are used.
- Do not pipette with mouth.



PROCEDURAL NOTES/LAB. QUALITY CONTROL

1. When not in use, kit components should be stored refrigerated as indicated on vials or bottles labels. All reagents should be warmed to room temperature before use. Lyophilized standards should be discarded directly after resuspension and use.
2. Once the desired number of strips has been removed, immediately reseal the bag to protect the remaining strips from deterioration.
3. Cover or cap all reagents when not in use.
4. Do not mix or interchange reagents between different batches.
5. Do not use reagents beyond the expiration date of the kit.
6. Use a clean disposable plastic pipette tip for each reagent, standard, or specimen addition in order to avoid cross-contamination; for the dispensing of H₂SO₄ and substrate solution, avoid pipettes with metal parts.
7. Use a clean plastic container to prepare the washing solution.
8. Thoroughly mix the reagents and samples before use by agitation or swirling.
9. All residual washing liquid must be drained from the wells by efficient aspiration or by decantation followed by tapping the plate forcefully on absorbent paper. Never insert absorbent paper directly into the wells.
10. The TMB solution is light sensitive. Avoid prolonged exposure to light. Also, avoid contact of the TMB solution with metal to prevent colour development. Warning TMB is toxic avoid direct contact with hands. Dispose of properly.
11. When pipetting reagents, maintain a consistent order of addition from well-to-well. This will ensure equal incubation times for all wells.
12. Respect incubation times described in the assay procedure.

SPECIMEN COLLECTION, PROCESSING AND STORAGE

Cell culture supernatants- Remove particulates and aggregates by spinning at approximately 1000 x g for 10 min.

Serum—Avoid any unintentional stimulation of the cells by the procedure. Use pyrogen/endotoxin free collecting tubes. Serum should be removed rapidly and carefully from the red cells after clotting. For that, after clotting, centrifuge at approximately 1000 x g for 10 min and remove serum.

Plasma-EDTA, citrate and heparin plasma can be assayed. Spin samples at 1000 x g for 30 min to remove particulates. Harvest plasma.

Storage-If not analyzed shortly after collection, samples should be aliquoted (250-500µl) to avoid freeze-thaw cycles and stored frozen at –70°C. Avoid multiple freeze-thaw cycles of frozen specimens.

When possible, avoid use of badly hemolyzed or lipemic sera. If large amounts of particles are present, they should be removed prior to assay by centrifugation or filtration.

Recommendation : Do not thaw by heating at 37°C or 56°C. Thaw at room temperature and make sure that sample is completely thawed and homogeneous before assaying

PREPARATION OF REAGENTS

Standard buffer diluent 10X concentrate

Dilute 10 times with distilled water before use.

Standards

Standard vials have to be reconstituted with the volume of standard diluent indicated on the vial. This reconstitution gives a stock solution of 25 pg/ml IFN γ . Allow standard to stand for 5 minutes with gentle swirling prior to making dilutions. Serial dilutions of standard must be made before each assay and cannot be stored.

Samples

Normal sera and plasmas may be applied undiluted. Nevertheless, sera or plasmas from patients with various pathologies may be applied undiluted and diluted (to prevent too high concentrations). Please note that certain sera/plasma may induce false positive (for example, in reason to presence of anti-mouse anti-IgG antibody). A simple sample dilution (1:2) allows eliminating interference. As IFN γ concentrations may vary considerably in cell supernatant samples, it is not easy to recommend a dilution factor.

Dilution of biotinylated anti-IFN γ

Dilute the biotinylated anti-IFN γ with the biotinylated antibody diluent in a clean glass vial according to the number of wells to be used. See the next table for volumes to pipette. Extemporaneous preparations are recommended.

Number of Wells	Biotinylated Antibody (µl)	Biotinylated Antibody Diluent (µl)
16	40	1060
32	80	2120
48	120	3180
96	240	6360



Dilutions of Streptavidin-HRP – Step L and R :



**Add 0.5 ml of HRP diluent to a 5 µl vial of Streptavidin-HRP
THIS PRE-DILUTION WILL BE USED FOR STEP L AND STEP R, BUT NOT FOR FUTHER EXPERIMENTS.**



From the previous solution two others dilutions must be prepared: one for the step “L” and one for the step “R”.

Extemporaneous preparations are recommended. Following the number of wells to be used, further dilutions of Streptavidin-HRP should be made with HRP diluent in a clean glass vial: see hereafter the table for volumes to pipette.

Number of Wells	Preparation of Streptavidin solution 1 – Step L		Preparation of Streptavidin solution 2 – Step R	
	Streptavidin-HRP (µl)	Strep-HRP Diluent (ml)	Streptavidin-HRP (µl)	Strep-HRP Diluent (ml)
16	20	1.980	40	1.960
32	40	3.960	80	3.920
48	60	5.940	120	5.880
96	120	11.880	240	11.760

Dilution of Amplifier

Extemporaneous preparations are recommended. Following the number of wells to be used, further dilutions of amplifier should be made with amplification diluent in a clean glass vial: see hereafter the table for volumes to pipette.

Number of Wells	Amplifier (µl)	Amplification Diluent (ml)
16	20	1.980
32	40	3.960
48	60	5.940
96	120	11.880

Washing Buffer 200X concentrate

Dilute 200 times in distilled water.

ASSAY METHOD

- a) Before use, mix all reagents thoroughly without making foam.
- b) Determine the number of microwell strips required to test the desired number of samples, plus appropriate number of wells needed for running blanks and standards. **Each sample, standard, blank should be assayed in duplicate.** Remove sufficient microwell strips from the pouch.
- c) Add 100 µl of standard diluent to standard wells B1, B2, C1, C2, D1, D2, E1, E2, F1, and F2. Reconstitute standard vial with the appropriate volume of standard diluent as described in the chapter reagents preparation. Pipet 200 µl of standard into wells A1 and A2 (see Plate Scheme below). Transfer 100 µl from A1 and A2 to B1 and B2 wells. Mix the contents by repeated aspirations and ejections. Take care not to scratch the inner surface of microwells. Repeat this procedure from the wells B1, B2 to wells C1, C2 and from wells C1, C2 to D1, D2 and so on creating two parallel rows of IFN γ standard dilutions ranging from 25 to 0.78 pg/ml. Discard 100 µl from the content of the last microwells used (F1, F2).
Alternatively these dilutions can be made in separate tubes and diluted standard pipetted directly into wells.
- d) Add 100 µl of standard diluent to the blank wells (G1-G2).
- e) Add 100 µl of sample to sample wells
- f) Cover with a plate cover and incubate for 1 hour at room temperature (18°C - 22°C) with slow shaking
- g) Remove the cover and wash the plate as follows:
 - 1) aspirate the liquid from each well ;
 - 2) dispense 0.3 ml of washing solution into each well ;
 - 3) aspirate again the content of each well ;
 - 4) Repeat steps 2) and 3) two times.
- h) Preparation of biotinylated anti-IFN γ : see reagents preparation.
- i) Add 50 µl of diluted biotinylated anti-IFN γ to all wells.
- j) Cover with a plate cover and incubate for 1 hour at room temperature (18°C - 22°C) with slow shaking
- k) Remove the cover and wash according to step g)
- l) Prepare streptavidin-HRP solution 1 just before use: see reagents preparation.
- m) Distribute 100 µl of the streptavidin-HRP solution 1 to all wells, including blank wells.
- n) Cover and incubate 20 min at room temperature with slow shaking.
- o) Remove the cover and wash according to step g)

All incubation steps
except substrate
TMB and acid are
performed under
slow shaking at
room temperature



- p) Add 100 µl of amplifier dilution prepared according the instructions for the preparation of reagents and incubate 15 minutes with slow shaking.
- q) Remove the cover and wash according to step g).
- r) Prepare streptavidin-HRP solution 2 just before use: see reagents preparation.
- s) Distribute 100 µl of the streptavidin-HRP solution 2 to all wells, including blank wells.
- t) Cover and incubate 20 min at room temperature with slow shaking.
- u) Remove the cover and wash according to step g).
- v) Pipette 100 µl of ready-to-use TMB substrate solution into all wells, including the blank wells and incubate in the dark within 5 minutes at room temperature. Avoid direct exposure to light by wrapping the plate in aluminium foil.
- w) Incubation time of the substrate solution is usually determined by the ELISA reader performances: many ELISA readers record absorbance only up to 2.0 O.D. **The O.D. values of the plate should be monitored and the substrate reaction stopped before positive wells are no longer properly readable.**
- x) The enzyme-substrate reaction is stopped by quickly pipetting 100 µl of stop reagent (1N sulphuric acid) into each well, including the blank wells, to completely and uniformly inactivate the enzyme. Results must be read rapidly after the addition of stop reagent.
- y) Read absorbance of each well on a spectrophotometer using 450 nm as the primary wavelength and optionally 620 nm (610 nm to 650 nm is acceptable) as the reference wavelength.

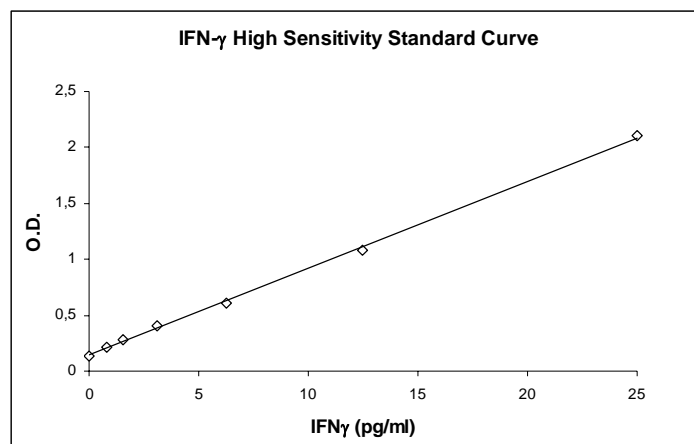
Note: In case of incubation without shaking the O.D values may be lower than with shaking; in this case let the color develop longer in order to obtain correct OD values

SUGGESTED PLATE SCHEME

	Standard Concentrations pg/mL		Sample wells									
	1	2	3	4	5	6	7	8	9	10	11	12
A	25	25										
B	12.5	12.5										
C	6.25	6.25										
D	3.12	3.12										
E	1.56	1.56										
F	0.78	0.78										
G	Blank	Blank										
H												

DATA ANALYSIS

Generate a linear standard curve by plotting the average absorbance on the vertical axis versus the corresponding IFN γ standard concentration on the horizontal axis. The amount of IFN γ in each sample is determined by extrapolating OD values to IFN γ concentrations using the standard curve.



Typical IFN γ High sensitivity standard curve ranging from 0.78 to 25 pg/mL



LIMITATIONS OF THE PROCEDURE

Do not extrapolate the standard curve beyond the 25 pg/ml standard curve point. The dose-response is non-linear in this region and therefore it is difficult to obtain accurate concentration. Concentrated samples (> 25 pg/ml) have to be diluted with standard diluent or with your own sample buffer. During analysis, multiply results by the appropriate dilution factor.

The influence of various drugs, aberrant sera (hemolyzed, hyperlipidemic, jaundiced...) has not been investigated. The rate of degradation of native IFN γ in various matrices has not been investigated.

PERFORMANCES AND CHARACTERISTICS

Sensitivity

This has been determined by adding 3 standard deviations to the mean concentration of 40 zeros. The minimum detectable dose of IFN γ was less than 0.69 pg/ml.

Specificity

Ten specificities were tested with concentrations higher than IFN γ curve concentrations. No cross reaction was observed for concentrations ranging from 2500 to 78.12 pg/ml for IL-1 α , IL-2, IL-8, IL-12p40, TNF- α , CD95 Fas, TRAIL, ICAM1, Gp130 et GM-CSF.

Spike - Recovery

The spike recovery was evaluated by spiking two levels of IFN γ into three human serum pools, two plasma pools and two culture media. Recovery was evaluated with one test. Mean recoveries were 92% in sera (range values: 84-98%), 72% in plasma (Range values: 61-81%) and 108% in cell culture media (range values: 88-117%).

Precision

Two human serum pools, two human plasma pools and two cell culture media samples with various concentrations of IFN γ were tested for repeatability and reproductibility. Each assay was carried out with 3 duplicates of each sample. Three independent assays were performed. The intra-assay and inter-assay coefficient of variation has been calculated to be 3.9% and 8.6% respectively.

Linearity of dilution

Two human serum pools, one human plasma pool and one cell culture medium samples with various concentrations of IFN γ were serially diluted in standard buffer diluent. Linearity was evaluated on 4 dilutions. The linear regression of samples versus the expected concentrations yielded a quote slope of 0.994.

Expected values

16 sera and 16 plasmas from healthy individual donors were tested undiluted and diluted in duplicates. Mean concentration in sera was 2.3 pg/ml (12 positives – range values of positive sera : 0.70 – 5.82 pg/ml) and in plasma is 1.89 pg/ml (13 positives – range values of positive plasmas : 0.89 – 5.08 pg/ml).

∇-----

Study : _____

Specificity : _____

Date : _____

Plate n°: _____

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												



ASSAY PROCEDURE SUMMARY:

Human High Sensitivity IFN γ ELISA Kit

Total procedure length: 3H00

Specific molecule detection steps	<p>Add 100μl of sample or diluted standard</p> <p style="text-align: center;">⇓</p> <p>Incubate 1 hour at room temperature with slow shaking</p> <p style="text-align: center;">⇓</p> <p>Wash three times</p> <p style="text-align: center;">⇓</p> <p>Add 50μl of diluted biotinylated detection antibody to all wells</p> <p style="text-align: center;">⇓</p> <p>Incubate 1 hour at room temperature with slow shaking</p>
Streptavidin –HRP / Amplification steps	<p style="text-align: center;">⇓</p> <p>Wash three times</p> <p style="text-align: center;">⇓</p> <p>Add 100μl of streptavidin-HRP (solution 1) to all wells</p> <p style="text-align: center;">⇓</p> <p>Incubate 20 min at room temperature with slow shaking</p> <p style="text-align: center;">⇓</p> <p>Wash three times</p> <p style="text-align: center;">⇓</p> <p>Add 100 μl amplifier to all wells</p> <p style="text-align: center;">⇓</p> <p>Incubate 15 min at room temperature with slow shaking</p> <p style="text-align: center;">⇓</p> <p>Wash three times</p> <p style="text-align: center;">⇓</p> <p>Add 100μl of streptavidin-HRP (solution 2) to all wells</p> <p style="text-align: center;">⇓</p> <p>Incubate 20 min at room temperature with slow shaking</p>
Revelation and reading steps	<p style="text-align: center;">⇓</p> <p>Wash three times</p> <p style="text-align: center;">⇓</p> <p>Add 100 μl of ready-to-use TMB Protect from light. Let the color develop for around 5 min.</p> <p style="text-align: center;">⇓</p> <p>Add 100 H₂SO₄</p> <p style="text-align: center;">⇓</p> <p>Read Absorbance at 450 nm</p>

